

# The Evaluation of Eight Commercially Available STR Kits

# Abstract:

The National Forensic Science Technology Center (NFSTC) conducted a study to evaluate the performance of eight STR amplification kits: Applied Biosystems' AmpfℓSTR® Profiler Plus® ID kit, Cofiler® kit, Identifiler® kit, MiniFiler™ kit and the Yfiler® kit and Promega's PowerPlex® 16 system, PowerPlex® Y system and the PowerPlex® 55 system.

The performance of a single source serial dilution was assessed with each STR amplification kit based on a defined set of criteria:

- Sensitivity
- Heterozygosity
- Baseline noise
- Reproducible amplification artifacts

Heterozygosity and sensitivity from two person mixture series were compared to results obtained from single source samples.

# Introduction:

There are various commercial STR multiplex kits available to the forensic science DNA community that are designed to address the ever-changing needs of crime laboratories. The primary goal of this study is to provide an overview of key performance measures of the eight STR kits tested.

# **Materials and Method:**

## Materials:

- Standards from 2 male donors
- Phenol:chloroform:Isoamyl alcohol (25:24:1)
- TE Buffer, DTT and Proteinase K (10ng/ul)
- Applied Biosystems' Human DNA Quantifiler® Kit
- Applied Biosystems' Ampf/STR<sup>®</sup> Profiler Plus<sup>®</sup> kit, Cofiler<sup>®</sup> kit, Identifiler<sup>®</sup> kit, MiniFiler<sup>™</sup> kit and the Yfiler<sup>®</sup>
- Promega's PowerPlex<sup>®</sup> 16 system, PowerPlex<sup>®</sup> Y system and the PowerPlex<sup>®</sup> S5 system
- Running Buffer, 10X
- 16 capillary array, 36cm
- POP-4<sup>™</sup> polymer for 3130*x*/
- Matrix standards
- Internal Lane Size Standards
- Hi-Di<sup>™</sup> Formamide
- 96-Well GeneAmp<sup>®</sup> PCR System 9700
- 7500 Real-Time PCR System
- 3130x/ Genetic Analyzer

## Method:

- Two separate known human male DNA standards were prepared utilizing a standard organic extraction method in conjunction with the Millipore Microcon<sup>®</sup> 100 centrifugal filter device.
- The samples were serially diluted to yield the following target concentrations: 1.0, 0.5, 0.25, 0.125, 0.0625, 0.03125, 0.015625, and 0.0078 ng.
- The two standards were systematically combined to create the following mixture ratios: 1:20, 1:15, 1:12, 1:10, 1:8, and 1:5. The targeted concentrations varied per kit and were based on the performance of the dilution series. (Profiler Plus<sup>®</sup>, Cofiler<sup>®</sup>, Identifiler<sup>®</sup> targeted 1 ng. PowerPlex<sup>®</sup> 16, PowerPlex<sup>®</sup> S5, PowerPlex<sup>®</sup> Y and Yfiler<sup>®</sup> targeted 0.5ng. MiniFiler<sup>™</sup> target was 0.25ng.)
- The samples were quantitated using the Applied Biosystems Quantifiler<sup>®</sup> Human Quantification Kit on an Applied Biosystems 7500 Real-Time PCR System. The results were normalized with NIST quantitation standards.
- Samples were amplified on an Applied Biosystems GeneAmp<sup>®</sup> PCR 9700 thermal cycler following manufacturer's specifications. Applied Biosystems' AmpfℓSTR<sup>®</sup> Profiler Plus<sup>®</sup> kit, Cofiler<sup>®</sup> kit, Identifiler<sup>®</sup> kit, MiniFiler<sup>™</sup> kit,

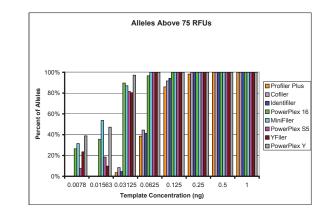
and the Yfiler<sup>®</sup> kit and Promega's PowerPlex<sup>®</sup> 16 system, PowerPlex<sup>®</sup> Y system, and the PowerPlex<sup>®</sup> S5 system.

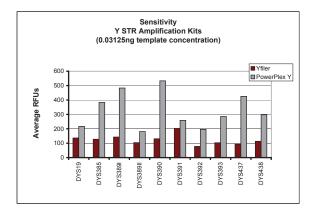
- The samples were then separated and detected using an Applied Biosystems 3130x/ Genetic Analyzer using manufacture's recommended conditions:
- Applied Biosystems kits: 3kv, 10 sec injections, 8.7  $\mu I$  Formamide, 0.3  $\mu I$  GS 500, 1  $\mu I$  of sample.
- Promega kits: 3kv, 10 sec injections, 9.5  $\mu I$  Formamide, 0.5  $\mu I$  ILS 600, 1  $\mu I$  of sample
- Data was analyzed using GeneMapper® ID Software v3.2.

# **Results and Discussion:**

#### Sensitivity:

- Concentrations at which alleles fell below 75 RFUs for each kit are depicted in table 1.
- MiniFiler<sup>™</sup> and PowerPlex<sup>®</sup> 16 produced the highest RFUs for each concentration.
- Identifiler<sup>®</sup>, Profiler Plus<sup>®</sup>, and Cofiler<sup>®</sup> exhibited comparable peak heights to each other for each concentration.
- Profiler Plus<sup>®</sup> displayed an RFU value less than 75 at 0.25 ng concentration at D7S820 in one injection of the triplicate data set.
- Note: Profiler Plus<sup>®</sup> and Cofiler<sup>®</sup> amplifications were performed with manufacture's recommended 50 µl reaction.
- The RFU range was higher for PowerPlex<sup>®</sup> Y than Yfiler<sup>®</sup> at each concentration.





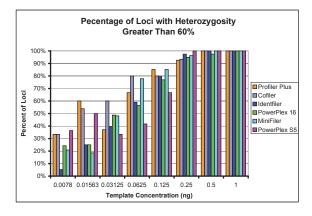
## Heterozygosity:

- As sample concentration decreases, the number of loci with heterozygosity greater than 60% also decreases.
- At concentrations of 0.5 ng and below, PowerPlex® 16 did not maintain a minimum heterozygosity of 60% at D8S1179

- 0.5 ng corresponds with an average RFU of 2200

- Note: this occurred once in the triplicate injection

- At concentrations of 0.25 ng and lower, Profiler Plus<sup>®</sup>, Cofiler<sup>®</sup>, Identifiler<sup>®</sup>, MiniFiler<sup>™</sup> did not maintain minimum of 60% heterozygosity.
- 0.25 ng concentration corresponds with the following peak height ranges:
- Profiler Plus<sup>®</sup> 138 to 273 RFUs
- Cofiler® 210 to 350 RFUs
- Identifiler  $^{\ensuremath{\text{\tiny B}}}$  255 to 445 RFUs
- MiniFiler™ 1870 to 3250 RFUs
- At concentrations of 0.125 ng and lower, PowerPlex<sup>®</sup> S5 displays loss of 60% heterozygosity.
- 0.125 ng corresponds with the RFU range 228 to 753



## **Amplification Artifacts:**

- Several dye blobs were present in the MiniFiler™, Yfiler<sup>®</sup>, Identifiler<sup>®</sup>, PowerPlex<sup>®</sup> 16, PowerPlex<sup>®</sup> Y, Profiler Plus<sup>®</sup> kits.
- Some instances of elevated stutter were observed for Profiler Plus<sup>®</sup>, Identifiler<sup>®</sup>, PowerPlex<sup>®</sup> 16.
- No elevated stutter was observed with Cofiler<sup>®</sup>, MiniFiler<sup>™</sup>, PowerPlex<sup>®</sup> S5.
- Indications of plus stutter were observed in several kits and in some instances were greater than 75 RFUs.
- Minus A was observed in several kits, most notably in PowerPlex® S5.

## **Baseline Noise:**

• All amplification kits displayed low background noise with the limit of detection (LOD) ranging from 10 to 15 RFUs.

 $LOD = \overline{X}_{blank} + 3\sigma_{blank}$ 

• The limit of quantitation (LOQ) ranging from 24 to 36 RFUs for all kits.

 $LOQ = \overline{X}_{blank} + 10\sigma_{blank}$ 

• In general, PowerPlex<sup>®</sup> 16 displayed a higher level of noise than the other kits tested.

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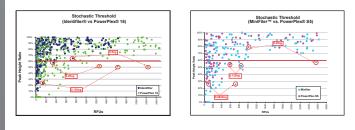


#### Mixtures:

	Profiler Plus®	Cofiler®	Identifiler®	PowerPlex® 16	PowerPlex® S5	MiniFiler™	Yfiler®	PowerPlex® Y
	Percentage of Minor Contributor Alleles Above 75 RFUs							
Mixture ratio								
1:5	100%	100%	100%	100%	100%	100%	100%	83%
1:8	100%	100%	100%	88%	100%	75%	87%	77%
1:10	82%	100%	85%	77%	100%	73%	93%	67%
1:12	59%	92%	74%	79%	94%	73%	87%	56%
1:15	54%	42%	68%	73%	94%	48%	78%	38%
1:20	18%	75%	55%	50%	81%	45%	69%	44%

# **Conclusions:**

- In general, the concentration at which heterozygosity fell below 60% was always higher than the concentration for which peak heights fell below 75 RFUs.
- PowerPlex<sup>®</sup> 16 appeared to be more sensitive than Identifiler<sup>®</sup>, Profiler Plus<sup>®</sup> and Cofiler<sup>®</sup>, however it displayed stochastic effects at higher RFU values/concentrations when compared to these kits.
- MiniFiler™ appeared to be more sensitive than PowerPlex<sup>®</sup> S5 however it displayed stochastic effects at higher RFU values/concentrations.
- Dropout (where a heterozygote appears as a homozygote) was observed with MiniFiler<sup>™</sup> and PowerPlex<sup>®</sup> 16 with approximate peak heights at 200 and 500 RFUs, respectively.
- Yfiler<sup>®</sup> and PowerPlex<sup>®</sup> Y perform similarly. Each kit had alleles that were below or less than 75 RFUs at 0.03125 ng.
- Laboratories should perform appropriate validation studies in order to establish interpretation guidelines which should include assessment of LOD, LOQ and stochastic threshold for each amplification kit and instrument.
- The mixture series performed as expected when compared to single source samples at comparable concentrations.



# **References:**

Internal Validation of STR Systems Reference Manual. www.promega.com part # GE053, revised 9/06.

The most current user and technical manuals for each kit were referenced in this study. Manuals procured from www.appliedbiosystems.com (published 1998-2006) and www.promega.com/tbs/ (published 2008.)

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1st concentration to fall below 60 % heterozygosity